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Mini Review

Manganese efflux in Parkinsonism: Insights from newly characterized *SLC30A10* mutations

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ABSTRACT

Although manganese (Mn) is required for normal cellular function, overexposure to this metal may cause an extrapyramidal syndrome resembling Parkinson's disease (PD). Notably, high whole-blood Mn levels have been reported in patients with idiopathic PD. Because Mn is both essential at low dose and toxic at higher dose; its transport and homeostasis are tightly regulated. Previously, the only protein known to be operant in cellular Mn export was the iron-regulating transporter, ferroportin (Fpn). The causal role for Mn in PD has yet to be fully understood, but evidence of a familial predisposition to PD associated with Mn toxicity is mounting. A recently discovered mutation in *SLC30A10* identified its gene product as putatively involved in Mn efflux. Patients with the *SLC30A10* mutation display Parkinsonian-like gate disturbances and hypermanganesemia. This review will address Mn transport proteins, the newly discovered *SLC30A10* mutations and their implications to Parkinsonism and Mn regulation.

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1. Introduction

Manganese (Mn) is an essential trace element, but at elevated levels it is also a risk factor for Parkinson's disease (PD). Mn homeostasis is required for a variety of cellular processes, and is necessary for normal central nervous system (CNS) functioning, including immune function, carbohydrate metabolism [1,2]. Under normal conditions, Mn functions as a cofactor for a variety of proteins including a subset of Mn-specific proteins, such as glutamate synthetase (GS) and superoxide dismutase 2 (SOD2, Mn-SOD) that are integral to cell detoxification and survival [3]. Mn uptake and distribution is important to normal cellular processes in the brain, including astrocyte morphology and migration, glutamate/ γ -aminobutyric acid (GABA) metabolism, and DAergic neuron maintenance [4–7].

Overexposure to Mn may be toxic, marked by accumulation of Mn in the basal ganglia as observed with magnetic resonance imaging (MRI) [8,9]. Overexposure may occur in occupational settings, especially from Mn mining dusts and welding fumes [10,11]. Welding fume-exposed workers are much more likely to experience Parkinsonism then matched controls [12]. Occupationally exposed workers have distinct MRI patterns with increased T1-weighted intensities in the basal ganglia, especially the globus pal-

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lidus [13]. In addition, exposures to Mn from medically administered parenteral nutrition have been reported [9,14,15], corroborating both inhalation and ingestion represent routes of absorption [16]. These overexposures may lead to manganism, defined as a Parkinsonism-like syndrome [17]. Notably, exposure to Mn has been implicated in idiopathic cases of Parkinsonism, including PD [18,19], and exposures from air or water are correlated with increased risk of PD [2,19,20]. The strongest correlation between any type of environmental exposure and increased PD risk is inherent to occupational exposures to Mn [16,19,21].

Parkinsonism encompasses a group of neurological disorders with the defining feature of marked motor impairment, presenting as tremor, postural instability and hypokinesia. PD is the leading cause of Parkinsonism and is the second leading cause of neurodegenerative diseases, affecting nearly 2% of persons above the age of 60 in the United States. Identified risk factors for PD include age, genetics, as well as environmental exposures. The risk of developing PD doubles every ten years after the age of 70 [22,23]. In the last two decades, 15 loci and 11 genes have been liked to familial inherited PD [24–26]. For nearly a century, we have understood that the progressive motor impairment in PD results from the loss of DAergic neurons in the midbrain substantia nigra (SN) [27], but definitive mechanisms for the etiology of PD are still elusive, with a majority of PD cases remaining idiopathic (of unknown origin).

Although idiopathic PD and Mn-induced Parkinsonism share motor impairment symptoms and DAergic cell loss, there are distinct differences between their clinical manifestations. Unlike

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idiopathic PD, manganism is more likely to present as dystonia without resting tremor, and in general manganism is less responsive to Levadopa therapy [8,17,28]. The average age of onset for Mn-induced parkinsonism after occupational exposures is 46, compared to 63 for idiopathic PD [29].

Genetic predisposition to Mn toxicity has been correlated with Parkin (PARK2) [30], a gene involved in autosomal recessive PD, and ATP13A2 (PARK9) [31,32], a putative Mn transporter. Parkin, most commonly mutated in autosomal recessive PD, encodes an E3 ubiquitin protein ligases thought to be involved in protein degradation and oxidative stress protection [26,33]. ATP13A2, mutated in less than five percent of autosomal recessive PD, encodes a cation transporter ATPase that transports Mn [31,34]. Patients carrying autosomal recessive mutations in Parkin and ATP13A2 also seem to be at increased risk for Mn toxicity [30,32,35]. In addition, patients with idiopathic PD have higher whole-blood Mn levels than individuals without the disease in the same population [36]. suggesting that brain Mn accumulation may be involved in the etiology of PD. In cases of PD and Mn overexposure, Mn accumulates in the brain regions associated with Parkinsonian motor dysfunction, primarily the globus pallidus and the substantia nigra of the basal ganglia [5].

Mn absorption, transport and excretion are tightly regulated. A number of proteins involved in Mn transport have been identified including the putative uptake proteins divalent metal transporter-1 (DMT1), transferrin receptor (TfR) and ATP13A2 (PARK9), as well as the efflux protein ferroportin (Fpn). For a more comprehensive review Mn transport please refer to Bowman et al. [1] and Au et al. [37].

Mutations in a putative Mn exporter gene *SLC30A10* have been recently described. These mutations are associated with marked motor impairment, including a Parkinsonism-like syndrome. The following discussion details known mechanisms of Mn transport, with a focus on the *SLC30A10* gene.

1.1. DMT1 and TfR: Fe and Mn absorption

Proteins involved in Mn transport share the characteristic of carrying more than a single metal species, including DMT1 and transferrin receptor (TfR). Dietary Fe and Mn are both absorbed through the intestine by DMT1 and possibly TfR [37–40]. Neurons express DMT1 and TfR for the uptake of both Fe and Mn [37,41,42]. DMT1 carries divalent Mn [2,37], whilst TfR carries trivalent Mn. A DMT1 haplotype has been identified as a risk factor for PD [38]; association between TfR polymorphism and PD has yet to be identified [43].

1.2. Ferroportin: Fe and Mn efflux

Ferroportin (Fpn), also known as solute carrier family 40 member 1 (SLC40A1) or iron (Fe)-regulated transporter, was originally described as an Fe exporter. Fpn is the only known protein for Fe-efflux [44]. Absorption, efflux, and distribution of Mn appear to be inversely related to stored Fe, with Fe deficiency facilitating Mn absorption [45-47]. Fpn is localized in all cells [41,44] and it was the first protein implicated in Mn-efflux [48-50]. Yin et al. identified Fpn as a Mn-transport protein, showing that Mn efflux decreases with decreased expression of Fpn [48,50]. Fpn is a transmembrane protein with 9-12 transmembrane domains [44] with both a cytosolic N and C terminus [51]. Fpn may function as a monomer or a dimer in Fe and Mn efflux [44,49]. Mutations in Fpn (SCL40A1) lead to Fe accumulation called haemochromatosis type 4, also known as Ferroportin disease [52,53]. No neurological symptoms are reported from Fpn mutation. The effect of SCL40A1 mutation on Mn accumulation remains unknown.

1.3. Fe in PD and Mn distribution

Fe and Mn share many features beyond transport and efflux proteins. Like Mn, Fe is also paradoxically a dietary requirement, as well as neurotoxin upon overexposure. Under normal conditions, Fe is required for heme oxygen transport throughout the body, as well as mitochondrial electron transport chain function [54–56]. Fe accumulation, analogous to Mn, has been implicated in PD [57–59]; thus an understanding of Fe transport has also become important in better characterizing the etiology of PD [43].

Of particular relevance to this review, Fe accumulation and distribution seems to be inversely related to Mn accumulation and distribution. Dietary Fe deficiency is one of the most common nutritional deficiencies, affecting nearly two million people world wide, unlike Mn deficiency, which is extremely rare [46,60,61]. Fe deficiency (anemia) can be a risk factor for Mn accumulation in the brain [60.62]. Fe deficiency increases expression of DMT1. TfR, and Fpn, where as Fe overexposure has been shown to decrease expression of DMT1 and Fpn [63-65]. In cases of hereditary haemochromatosis, patients experience Fe overload and hepatic cirrhosis, and DMT1 and Fpn expression is uncoupled from the body Fe status [65]. Fe deficiency increases blood and brain Mn levels in exposed humans and animals [66-68], whereas repletion with dietary Fe reduces Mn accumulation [68,69]. Furthermore, Mn overexposure promotes release of Fe stores and reduces cellular Fe load [70,71]. Taking into account the prevalence of global Fe deficiency (see above), this clinical condition represents a risk factor for increased Mn accumulation.

1.4. SLC30A10 and Mn efflux

Recently, the first example of a familial inherited mutation in Mn metabolism was observed in 10 different families [69,72,73]. Three papers described a disorder of Mn metabolism first identified by Tuschl et al. [74], with symptoms including hypermanganesemia, dystonia, polycythemia, and often, hepatic cirrhosis.

The cause of flawed Mn metabolism was identified as an autosomal recessive mutation in *SLC30A10* [69]. Tuschl et al. 2012 described eight families with non-symptomatic individuals with heterozygous *SLC30A10* profiles, and symptomatic individuals with homozygous *SLC30A10* mutations that appear to missense or truncation mutations that render a nonexistent or non-functional protein product. Symptom onset for *SLC30A10* mutations range between 2 and 57 years of age, with most cases presenting in childhood [69,72,73], suggesting a diverse environmental or multi-genetic component to the disease.

SLC30A10 was originally thought to be a zinc (Zn) transporter based on sequence analysis, but upon closer examination, SLC30A10 was identified as an Mn efflux protein. SLC30A10 belongs to the cation diffusion facilitator superfamily of metal transporters responsible for transport of Fe, Cu, Zn, and Mn. SLC30A10 is a 485 amino acid membrane embedded protein with 6 purposed transmembrane domains (TMDs) and a cytoplasmic N and C-termini [73]. Quadi et al. made the argument for SLC30A10 as a Mn transporter based on its amino acid sequence, noting that it contains a conserved NxxxD motif replacing the histidine-rich region characteristic of zinc (Zn) transporters and contains a conserved cytosolic cysteine at TMD IV, similar to other Mn specific transporters. Further, Tuschl et al. demonstrated that Mn sensitive yeast can be rescued by expression of human SLC30A10, but not mutated human SLC30A10, establishing SLC30A10 as an Mn transporter. SLC30A10 is expressed in the liver and neuronal cells of the globus pallidus under normal conditions, and is absent from the liver upon SLC30A10 mutation. Additionally, SLC30A10 is expressed in the retina and SLC30A10 is located on chromosome 1q41 near loci thought to influence high myopia.

The hypermanganesemia associated with *SLC30A10* mutation is extreme, with patients having whole blood Mn levels of 1200–6400 nmol/L, compared with normal whole blood Mn (<320 nmol/L). Interestingly, patients with *SLC30A10* mutation also show low Fe and increased total Fe binding capacity (TIBC), suggestive of Fe deficiency [69]. Along with high circulating levels, Mn appears to accumulate in the brains of patients with *SLC30A10* mutations. MRI showed T1-weighted intensities in the basal ganglia, as well as the cerebellum and anterior pituitary [72,73]. This MRI pattern of Mn accumulation is analogous to those seen in occupational exposures to high levels of Mn in welding fumes [2,10,21].

Dystonia or Parkinsonism is found in all cases with *SLC30A10* mutation. Patients with childhood onset of symptoms show primary dystonia, similar to very early onset of PD, where dystonia is the commonly observed motor symptom [26,75]. Childhood vs. adult onset differences in neuromotor disorders have been explained by basal ganglia maturity, and could explain the differences in motor symptoms seen in *SLC30A10* mutation. Treatment for patients with *SLC30A10* mutation involved oral Fe supplementation to increase the competition between Mn and Fe at shared transporters, such as DMT1 and TfR. After a month of Fe supplementation, patients showed marked motor improvement.

The pioneer studies on *SLC30A10* mutations have shed new light on the understanding of its product in Mn-induced neurotoxicity. However, additional research is required to better characterize the role of SLC30A10 in Mn homeostasis. To date, the subcellular localization of this protein has yet to be identified. Absent this knowledge, the mechanism(s) of SLC30A10 regulated Mn export will be hard to decipher. Does it directly export Mn from the cytosol to the extracellular matrix or to organelles, such as the mitochondria or Golgi for later excretion? Moreover, as Fe supplementation alleviates the symptoms caused by *SLC30A10* mutations, does SLC30A10 specifically regulate Mn as well as Fe efflux? Studies addressing these questions will facilitate the understanding on the role of SLC30A10 in preventing Mn accumulation and its parkinsonism-like effect.

In summary, SLC30A10 mutations highlight a new Mn metabolic disorder and offer a new chance to understand the role of Mn in Parkinsonism. Previously, occupational and environmental exposure to Mn has shown neurological symptoms of Parkinsonism, and Mn accumulation in the basal ganglia. SLC30A10 mutations are the first example of a genetic cause correlating neurological symptoms directly with Mn accumulation in humans. Symptoms of SLC30A10 mutations, including Parkinsonism, are improved with oral Fe supplementation, adding weight to the case for Fe deficiency as a risk factor for Mn-accumulation associated Parkinsonism. It is possible that heterozygous mutations or single nucleotide mutations in SLC30A10 may interact with environmental or occupational exposure to promote and exacerbate the onset of parkinsonism that would have previously been identified as idiopathic. Further research into the role of SLC30A10 and other potential putative Mn exporters is needed to determine its possible role in broader cases of Mn exposure and Parkinsonism.

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References

 A.B. Bowman, G.F. Kwakye, E.H. Hernández, M. Aschner, Role of manganese in neurodegenerative diseases, J. Trace Elem. Med. Biol. 25 (2011) 191–203.

- [2] M. Aschner, K.M. Erikson, E.H. Hernández, R. Tjalkens, Manganese and its role in disease: from transport to neuropathology, Neuromolecular Med. 11 (2009) 252–266.
- [3] M. Morello, P. Zatta, P. Zambenedetti, A. Martorana, V. D'Angelo, G. Melchiorri, G. Bernardi, G. Sancesario, Manganese intoxication decreases the expression of manganoproteins in the rat basal ganglia: an immunohistochemical study, Brain Res. Bull. 74 (2007) 406–415.
- [4] M. Sidoryk-Wegrzynowicz, E. Lee, J. Albrecht, M. Aschner, Manganese disrupts astrocyte glutamine transporter expression and function, J. Neurochem. 110 (2009) 822–830.
- [5] G.D. Stanwood, D.B. Leitch, V. Savchenko, J. Wu, V.A. Fitsanakis, D.J. Anderson, J.N. Stankowski, M. Aschner, B. McLaughlin, Manganese exposure is cytotoxic and alters dopaminergic and GABAergic neurons within the basal ganglia, J. Neurochem. 110 (2009) 378–389.
- [6] C. Kern, D. Smith, Preweaning Mn exposure leads to prolonged astrocyte activation and lasting effects on the dopaminergic system in adult male rats, Synapse 65 (2010) 532–544.
- [7] H. Yokoyama, H. Uchida, H. Kuroiwa, J. Kasahara, T. Araki, Role of glial cells in neurotoxin-induced animal models of Parkinson's disease, Neurol. Sci. 32 (2011) 1–7.
- [8] Y. Kim, J.W. Kim, K. Ito, H.S. Lim, H.K. Cheong, J.Y. Kim, Y.C. Shin, K.S. Kim, Y. Moon, Idiopathic parkinsonism with superimposed manganese exposure: utility of positron emission tomography, Neurotoxicology 20 (1999) 249–252.
- [9] M.G. Cersosimo, W.C. Koller, The diagnosis of manganese-induced parkinsonism, Neurotoxicology 27 (2006) 340–346.
- [10] B.A. Racette, L. McGee-Minnich, S.M. Moerlein, J.W. Mink, T.O. Videen, J.S. Perlmutter, Welding-related parkinsonism: clinical features, treatment, and pathophysiology, Neurology 56 (2001) 8–13.
- [11] K. Sriram, G.X. Lin, A.M. Jefferson, J.R. Roberts, R.S. Chapman, B.T. Chen, J.M. Soukup, A.J. Ghio, J.M. Antonini, Dopaminergic neurotoxicity following pulmonary exposure to manganese-containing welding fumes, Arch. Toxicol. 84 (2010) 521–540.
- [12] B.A. Racette, S.R. Criswell, J.I. Lundin, A. Hobson, N. Seixas, P.T. Kotzbauer, B.A. Evanoff, J.S. Perlmutter, J. Zhang, L. Sheppard, H. Checkoway, Increased risk of parkinsonism associated with welding exposure, Neurotoxicology 33 (2012) 1356–1361.
- [13] S.R. Criswell, J.S. Perlmutter, J.L. Huang, N. Golchin, H.P. Flores, A. Hobson, M. Aschner, K.M. Erikson, H. Checkoway, B.A. Racette, Basal ganglia intensity indices and diffusion weighted imaging in manganese-exposed welders, Occup. Environ. Med. 69 (2012) 437–443.
- [14] R.N. Dickerson, Manganese intoxication and parenteral nutrition, Nutrition 17 (2001) 689–693.
- [15] J.A. Chalela, L. Bonillha, R. Neyens, A. Hays, Manganese encephalopathy: an under-recognized condition in the intensive care unit, Neurocrit. Care 14 (2010) 456–458.
- [16] L. Normandin, M. Panisset, J. Zayed, Manganese neurotoxicity: behavioral, pathological, and biochemical effects following various routes of exposure, Rev. Environ. Health 17 (2002) 189–217.
- [17] D.B. Calne, N.S. Chu, C.C. Huang, C.S. Lu, W. Olanow, Manganism and idiopathic parkinsonism: similarities and differences, Neurology 44 (1994) 1583–1586.
- [18] J.M. Gorell, B.A. Rybicki, C. Cole Johnson, E.L. Peterson, Occupational metal exposures and the risk of Parkinson's disease, Neuroepidemiology 18 (1999) 303-308
- [19] H.K. Hudnell, Effects from environmental Mn exposures: a review of the evidence from non-occupational exposure studies, Neurotoxicology 20 (1999) 379–397.
- [20] M. Finkelstein, M. Jerrett, A study of the relationships between Parkinson's disease and markers of traffic-derived and environmental manganese air pollution in two canadian cities, Environ. Res. 104 (2007) 420–432.
- [21] J.M. Gorell, C.C. Johnson, B.A. Rybicki, E.L. Peterson, G.X. Kortsha, G.G. Brown, R.J. Richardson, Occupational exposure to manganese, copper, lead, iron, mercury and zinc and the risk of Parkinson's disease, Neurotoxicology 20 (1999) 239–247.
- [22] I.S. Pienaar, J. Götz, M.B. Feany, Parkinson's disease: insights from non-traditional model organisms, Prog. Neurobiol. 92 (2010) 558–571.
- [23] L.S. Forno, Neuropathology of Parkinson's disease, J. Neuropathol. Exp. Neurol. 55 (1996) 259–272.
- [24] K.R. Kumar, K. Lohmann, C. Klein, Genetics of Parkinson disease and other movement disorders, Curr. Opin. Neurol. 25 (2012) 466–474.
- [25] K. Nuytemans, J. Theuns, M. Cruts, C. Van Broeckhoven, Genetic etiology of Parkinson disease associated with mutations in the SNCA, PARK2, PINK1, PARK7, and LRRK2 genes: a mutation update, Hum. Mutat. 31 (2010) 763–780.
- [26] V. Bonifati, Genetics of Parkinson's disease, Minerva Med. 96 (2005) 175–186.
 [27] A.J. Lees, M. Selikhova, L.A. Andrade, C. Duyckaerts, The black stuff and Konstantin Nikolaevich Tretiakoff, Mov. Disord. 23 (2008) 777–783.
- [28] C.S. Lu, C.C. Huang, N.S. Chu, D.B. Calne, Levodopa failure in chronic manganism, Neurology 44 (1994) 1600–1602.
- [29] K. Erikson, M. Aschner, Increased manganese uptake by primary astrocyte cultures with altered iron status is mediated primarily by divalent metal transporter, Neurotoxicology 27 (2006) 125–130.
- [30] A.A. Aboud, A.M. Tidball, K.K. Kumar, M.D. Neely, K.C. Ess, K.M. Erikson, A.B. Bowman, Genetic risk for Parkinson's disease correlates with alterations in neuronal manganese sensitivity between two human subjects, Neurotoxicology (2012) 1–7.
- [31] A. Gitler, A. Chesi, M. Geddie, K. Strathearn, S. Hamamichi, K. Hill, K. Caldwell, G. Caldwell, A. Cooper, J. Rochet, S. Lindquist, Alpha-synuclein is part of a

- diverse and highly conserved interaction network that includes PARK9 and manganese toxicity, Nat. Genet. 41 (2009) 308–315.
- [32] J. Tan, T. Zhang, L. Jiang, J. Chi, D. Hu, Q. Pan, D. Wang, Z. Zhang, Regulation of intracellular manganese homeostasis by Kufor-Rakeb syndrome-associated ATP13A2 protein, J. Biol. Chem. 286 (2011) 29654–29662.
- [33] H. Yang, H.Y. Zhou, B. Li, G.Z. Niu, S.D. Chen, Downregulation of parkin damages antioxidant defenses and enhances proteasome inhibition-induced toxicity in PC12 cells, J. Neuroimmune pharmacol. 2 (2007) 276–283.
- [34] S.A. Schneider, K.P. Bhatia, Rare causes of dystonia parkinsonism, Curr. Neurol. Neurosci. Rep. 10 (2010) 431–439.
- [35] J.A. Roth, S. Singleton, J. Feng, M. Garrick, P.N. Paradkar, Parkin regulates metal transport via proteasomal degradation of the 1B isoforms of divalent metal transporter 1, J. Neurochem. 113 (2010) 454–464.
- [36] T. Fukushima, X. Tan, Y. Luo, H. Kanda, Relationship between blood levels of heavy metals and Parkinson's disease in china, Neuroepidemiology 34 (2010) 18–24.
- [37] C. Au, A. Benedetto, M. Aschner, Manganese transport in eukaryotes: the role of DMT1, Neurotoxicology 29 (2008) 569–576.
- [38] Q. He, T. Du, X. Yu, A. Xie, N. Song, Q. Kang, J. Yu, L. Tan, J. Xie, H. Jiang, DMT1 polymorphism and risk of Parkinson's disease, Neurosci. Lett. 501 (2011) 128– 131.
- [39] R. Settivari, J. Levora, R. Nass, The divalent metal transporter homologues SMF-1/2 mediate dopamine neuron sensitivity in caenorhabditis elegans models of manganism and parkinson disease, J. Biol. Chem. 284 (2009) 35758–35768.
- [40] M. Aschner, T.R. Guilarte, U. Dydak, S.R. Criswell, W. Zheng, Neurotoxicology 33 (2012) 881–886, http://dx.doi.org/10.1016/j.neuro.2011.12.010.
- [41] T. Moos, T. Rosengren Nielsen, Ferroportin in the postnatal rat brain: implications for axonal transport and neuronal export of iron, Semin. Pediatr. Neurol. 13 (2006) 149–157.
- [42] A. Benedetto, C. Au, M. Aschner, Manganese-induced dopaminergic neurodegeneration: insights into mechanisms and genetics shared with Parkinson's disease, Chem. Rev. 109 (2009) 4862–4884.
- [43] V. Greco, E.V. De Marco, F.E. Rocca, F. Annesi, D. Civitelli, G. Provenzano, P. Tarantino, V. Scornaienchi, F. Pucci, M. Salsone, F. Novellino, M. Morelli, S. Paglionico, A. Gambardella, A. Quattrone, G. Annesi, Association study between four polymorphisms in the HFE, TF and TFR genes and Parkinson's disease in southern italy, Neurol. Sci. 32 (2011) 525–527.
- [44] D.M. Ward, J. Kaplan, Ferroportin-mediated iron transport: expression and regulation, BBA-Mol. Cell Res. 2012 (1823) 1426–1433.
- [45] K.M. Erikson, Z.K. Shihabi, J.L. Aschner, M. Aschner, Manganese accumulates in iron-deficient rat brain regions in a heterogeneous fashion and is associated with neurochemical alterations, Biol Trace Elem. Res. 87 (2002) 143–156.
- [46] V.A. Fitsanakis, N. Zhang, S. Garcia, M. Aschner, Manganese (Mn) and iron (Fe): interdependency of transport and regulation, Neurotox. Res. 18 (2009) 124– 131.
- [47] X. Wang, G.J. Li, W. Zheng, Efflux of iron from the cerebrospinal fluid to the blood at the blood-csf barrier: effect of manganese exposure, Exp. Biol. Med. 233 (2008) 1561–1571.
- [48] Z. Yin, H. Jiang, E.-S.Y. Lee, M. Ni, K.M. Erikson, D. Milatovic, A.B. Bowman, M. Aschner, Ferroportin is a manganese-responsive protein that decreases manganese cytotoxicity and accumulation, J. Neurochem. 112 (2010) 1190–1198.
- [49] I. De Domenico, D.M. Ward, G. Musci, J. Kaplan, Evidence for the multimeric structure of ferroportin, Blood 109 (2007) 2205–2209.
- [50] M.S. Madejczyk, N. Ballatori, The iron transporter ferroportin can also function as a manganese exporter, BBA-Biomembranes 2012 (1818) 651–657.
- [51] X.-B. Liu, F. Yang, D.J. Haile, Functional consequences of ferroportin 1 mutations, Blood Cells Mol. Dis. 35 (2005) 33–46.
- [52] R. Mayr, W.J.H. Griffiths, M. Hermann, I. McFarlane, D.J. Halsall, A. Finkenstedt, A. Douds, S.E. Davies, A.R. Janecke, W. Vogel, T.M. Cox, H. Zoller, Identification of mutations in SLC40A1 that affect ferroportin function and phenotype of human ferroportin iron overload, Gastroenterology 140 (2011) 2056–2063. e2051.
- [53] M. Speletas, A. Kioumi, G. Loules, P. Hytiroglou, J. Tsitouridis, J. Christakis, A.E. Germenis, Analysis of SLC40A1 gene at the mRNA level reveals rapidly the causative mutations in patients with hereditary hemochromatosis type IV, Blood Cells Mol. Dis. 40 (2008) 353–359.

- [54] M. Hoppe, B. Brün, M.P. Larsson, L. Moraeus, L. Hulthén, Heme iron-based dietary intervention for improvement of iron status in young women, Nutrition 29 (2013) 89–95.
- [55] R.C. Hider, X. Kong, Iron speciation in the cytosol: an overview, Dalton Tras. (2012), [Epub ahead of print].
- [56] V.A. Fitsanakis, N. Zhang, S. Garcia, M. Aschner, Manganese (Mn) and iron (Fe): interdependency of transport and regulation, Neurotox. Res. 18 (2010) 124– 131.
- [57] H. Mochizuki, T. Yasuda, Iron accumulation in Parkinson's disease, J. Neural. Transm. 119 (2012) 1511–1514.
- [58] T. Moos, T.R. Nielsen, T. Skjørringe, E.H. Morgan, Iron trafficking inside the brain, J. Neurochem. 103 (2007) 1730–1740.
- [59] D.T. Dexter, F.R. Wells, A.J. Lees, F. Agid, Y. Agid, P. Jenner, C.D. Marsden, Increased nigral iron content and alterations in other metal ions occurring in brain in Parkinson's disease, J. Neurochem. 52 (1989) 1830–1836.
- [60] K.M. Erikson, T. Syversen, J.L. Aschner, M. Aschner, Interactions between excessive manganese exposures and dietary iron-deficiency in neurodegeneration, Environ. Toxicol. pharmacol. 19 (2005) 415–421.
- [61] J. Beard, K. Erikson, B.C. Jones, Neonatal iron deficiency results in irreversible changes in dopamine function in rats, J. Nutr. 133 (2003) 1174–1179.
- [62] M. Aschner, G. Shanker, K. Erikson, J. Yang, L.A. Mutkus, The uptake of manganese in brain endothelial cultures, Neurotoxicology 23 (2002) 165–168.
- [63] K.M. Erikson, B.C. Jones, J.L. Beard, Iron deficiency alters dopamine transporter functioning in rat striatum, J. Nutr. 130 (2000) 2831–2837.
- [64] K.M. Erikson, D.J. Pinero, J.R. Connor, J.L. Beard, Regional brain iron, ferritin and transferrin concentrations during iron deficiency and iron repletion in developing rats, J. Nutr. 127 (1997) 2030–2038.
- [65] H. Zoller, R.O. Koch, I. Theurl, P. Obrist, A. Pietrangelo, G. Montosi, D.J. Haile, W. Vogel, G. Weiss, Expression of the duodenal iron transporters divalent-metal transporter 1 and ferroportin 1 in iron deficiency and iron overload, Gastroenterology 120 (2001) 1412–1419.
- [66] H. Nam, M.D. Knutson, Effect of dietary iron deficiency and overload on the expression of ZIP metal-ion transporters in rat liver, Biometals 25 (2012) 115– 124
- [67] P. Brna, K. Gordon, J.M. Dooley, V. Price, Manganese toxicity in a child with iron deficiency and polycythemia, J. Child Neurol. 26 (2011) 891–894.
- [68] Y. Kim, B.K. Lee, Iron deficiency increases blood manganese level in the korean general population according to KNHANES 2008, Neurotoxicology 32 (2011) 247–254
- [69] K. Tuschl, P.T. Clayton, S.M. Gospe Jr, S. Gulab, S. Ibrahim, P. Singhi, R. Aulakh, R.T. Ribeiro, O.G. Barsottini, M.S. Zaki, M.L. Del Rosario, S. Dyack, V. Price, A. Rideout, K. Gordon, R.A. Wevers, W.K.K. Chong, P.B. Mills, Syndrome of hepatic cirrhosis, dystonia, polycythemia, and hypermanganesemia caused by mutations in SLC30A10, a manganese transporter in man, Am. J. Hum. Genet. 90 (2012) 457–466.
- [70] C. Kwik-Uribe, D. Smith, Temporal responses in the disruption of iron regulation by manganese, J. Neurosci. Res. 83 (2006) 1601–1610.
- [71] D.R. Crooks, M.C. Ghosh, M. Braun-Sommargren, T.A. Rouault, D. Smith, Manganese targets m-aconitase and activates iron regulatory protein 2 in AF5 GABAergic cells, J. Neurosci. Res. 85 (2007) 1797–1809.
- [72] M. Stamelou, K. Tuschl, W.K. Chong, A.K. Burroughs, P.B. Mills, K.P. Bhatia, P.T. Clayton, Dystonia with brain manganese accumulation resulting from SLC30A10mutations: a new treatable disorder, Mov. Disord. 27 (2012) 1317–1322.
- [73] M. Quadri, A. Federico, T. Zhao, G.J. Breedveld, C. Battisti, C. Delnooz, L.-A. Severijnen, L. Di Toro Mammarella, A. Mignarri, L. Monti, A. Sanna, P. Lu, F. Punzo, G. Cossu, R. Willemsen, F. Rasi, B.A. Oostra, B.P. van de Warrenburg, V. Bonifati, Mutations in SLC30A10 cause Parkinsonism and dystonia with hypermanganesemia, polycythemia, and chronic liver disease, Am. J. Hum. Genet. 90 (2012) 467–477.
- [74] K. Tuschl, P.B. Mills, H. Parsons, M. Malone, D. Fowler, M. Bitner-Glindzicz, P.T. Clayton, Hepatic cirrhosis, dystonia, polycythaemia and hypermanganesaemia-a new metabolic disorder, J. Inherit. Metab. Dis. 31 (2008) 151–163.
- [75] N.Z. Baquer, A. Taha, P. Kumar, P. McLean, S.M. Cowsik, R.K. Kale, R. Singh, D. Sharma, A metabolic and functional overview of brain aging linked to neurological disorders, Biogerontology 10 (2009) 377–413.